

Tech File: Hydrazine

AFAMRL-TR-84-035

ADA143122

ulation



**TOXICOKINETICS OF HYDRAZINE ADMINISTERED
PERCUTANEOUSLY TO THE RABBIT.**

W. C. KELLER
J. P. F. MURPHY
R. H. BRUNER
M. E. ANDERSEN
C. T. OLSON

JUNE 1984

20060707088

Approved for public release; distribution unlimited.

AIR FORCE AEROSPACE MEDICAL RESEARCH LABORATORY
AEROSPACE MEDICAL DIVISION
AIR FORCE SYSTEMS COMMAND
WRIGHT-PATTERSON AIR FORCE BASE, OHIO

STINFO COPY

NOTICES

When US Government drawings, specifications, or other data are used for any purpose other than a definitely related Government procurement operation, the Government thereby incurs no responsibility nor any obligation whatsoever, and the fact that the Government may have formulated, furnished, or in any way supplied the said drawings, specifications, or other data, is not to be regarded by implication or otherwise, as in any manner licensing the holder or any other person or corporation, or conveying any rights or permission to manufacture, use, or sell any patented invention that may in any way be related thereto.

Please do not request copies of this report from Air Force Aerospace Medical Research Laboratory. Additional copies may be purchased from:

National Technical Information Service
5285 Port Royal Road
Springfield, Virginia 22161

Federal Government agencies and their contractors registered with Defense Technical Information Center should direct requests for copies of this report to:

Defense Technical Information Center
Cameron Station
Alexandria, Virginia 22314

TECHNICAL REVIEW AND APPROVAL

AFAMRL-TR-84-035

The experiments reported herein were conducted according to the "Guide for the Care and Use of Laboratory Animals," Institute of Laboratory Animal Resources, National Research Council.

This report has been reviewed by the Office of Public Affairs (PA) and is releasable to the National Technical Information Service (NTIS). At NTIS, it will be available to the general public, including foreign nations.

This technical report has been reviewed and is approved for publication.

FOR THE COMMANDER



BRUCE O. STUART, PhD
Director Toxic Hazards Division
Air Force Aerospace Medical Research Laboratory

REPORT DOCUMENTATION PAGE

1a. REPORT SECURITY CLASSIFICATION			1b. RESTRICTIVE MARKINGS		
2a. SECURITY CLASSIFICATION AUTHORITY			3. DISTRIBUTION/AVAILABILITY OF REPORT		
2b. DECLASSIFICATION/DOWNGRADING SCHEDULE			Approved for public release; distribution unlimited.		
4. PERFORMING ORGANIZATION REPORT NUMBER(S)			5. MONITORING ORGANIZATION REPORT NUMBER(S)		
AFAMRL-TR-84-035					
6a. NAME OF PERFORMING ORGANIZATION Air Force Aerospace Medical Research Laboratory, Air Force Systems Command		6b. OFFICE SYMBOL (If applicable) THT	7a. NAME OF MONITORING ORGANIZATION		
6c. ADDRESS (City, State and ZIP Code) AFAMRL/THT Building 79, Area B Wright-Patterson AFB OH 45433			7b. ADDRESS (City, State and ZIP Code)		
8a. NAME OF FUNDING/SPONSORING ORGANIZATION		8b. OFFICE SYMBOL (If applicable)	9. PROCUREMENT INSTRUMENT IDENTIFICATION NUMBER		
8c. ADDRESS (City, State and ZIP Code)			10. SOURCE OF FUNDING NOS.		
			PROGRAM ELEMENT NO. 62202F	PROJECT NO. 6302	TASK NO. 08
11. TITLE (Include Security Classification) (U) Toxicokinetics of Hydrazine Administered Percutaneously to the Rabbit.			WORK UNIT NO. 04		
12. PERSONAL AUTHOR(S) W. C. Keller, J. P. F. Murphy, R. H. Bruner, M. E. Andersen, and C. T. Olson					
13a. TYPE OF REPORT FINAL		13b. TIME COVERED FROM Oct 80 TO Dec 82	14. DATE OF REPORT (Yr., Mo., Day) 84/06/13		15. PAGE COUNT 13
16. SUPPLEMENTARY NOTATION					
17. COSATI CODES			18. SUBJECT TERMS (Continue on reverse if necessary and identify by block number)		
FIELD	GROUP	SUB. GR.			
			Hydrazine Percutaneous Toxicokinetics		
			Amerzine H70		
19. ABSTRACT (Continue on reverse if necessary and identify by block number)					
<p>The percutaneous absorption of various aqueous concentrations of hydrazine over selected time periods has been studied. Duration of hydrazine exposure and hydrazine concentration correlated closely with the percent dose absorbed and peak serum hydrazine concentration. The data suggested that about 70% of the available hydrazine in H70 is still unabsorbed two minutes following hydrazine application. There was an increase in serum hydrazine concentration following removal of hydrazine from the skin surface suggesting the existence of an epidermal compartment.</p>					
20. DISTRIBUTION/AVAILABILITY OF ABSTRACT			21. ABSTRACT SECURITY CLASSIFICATION		
UNCLASSIFIED/UNLIMITED <input type="checkbox"/> SAME AS RPT. <input checked="" type="checkbox"/> DTIC USERS <input type="checkbox"/>					
22a. NAME OF RESPONSIBLE INDIVIDUAL		22b. TELEPHONE NUMBER (Include Area Code)		22c. OFFICE SYMBOL	
Lt Col Carl T. Olson		(513) 255-3364		AFAMRL/THT	

Preface

This research was performed in the Toxicology Branch, Toxic Hazards Division, Air Force Aerospace Medical Research Laboratory from October 1980 through December 1982. It was performed in support of Project 6302, "Occupational and Environmental Toxic Hazards in Air Force Operations"; Task 630208, "Toxicology of Aerospace Fuels"; Work Unit 63020804, "Chronic Toxicology of Hydrazine Strategic Missile Fuels".

The authors acknowledge the technical assistance of SSgt Temistocles Whittaker, TSGT Ernest Fairbrother, and Ms. Mary Buchwalder.

INTRODUCTION

Hydrazine, H_2NNH_2 , a polar, hydrophilic base, is a strong reducing agent with wide application in a variety of industrial processes. It is estimated that 90,000 workers are potentially exposed to hydrazine (Hz) or its salts each year (NIOSH, 1978). Hydrazine has been used in military and aerospace related systems as a propellant for many years. It was originally used as a jet fuel and a torpedo propellant during World War II in Germany, and more recently has been used in Titan II missiles, the emergency power unit (EPU) of the F-16, and the auxiliary power units of the space shuttle. The possibility that AF aircraft maintenance personnel may be subject to accidental Hz skin exposure is substantial. In addition, the proposed use of catalyzed aqueous solutions of Hz as corrosion inhibitors in naval propulsion boilers raises the possibility that Navy personnel may also be exposed to Hz.

Some data are available regarding skin exposure to Hz. The percutaneous absorption of Hz was described by Smith and Clark (1972) who applied Hz to the skin of anesthetized dogs at doses of 3-15 mmole/kg. A chemical burn occurred at the site of application, and absorption appeared complete by 70 minutes in all dose groups and sooner in the lower dose groups. The high dose group (15 mmole/kg) had a peak serum concentration of 60 $\mu\text{g/ml}$ while the low dose group had a maximum serum concentration of 2 $\mu\text{g/ml}$. The dermal LD_{50} for the dog was established as 90 mg/kg; for the rabbit, the dermal LD_{50} is 91 mg/kg, as compared to an iv LD_{50} of 20 mg/kg (Rotherberg and Cope, 1955). More recent research has been done concerning the percutaneous absorption of Hz in the rabbit (Keller et al., 1981) and this report relates the results of the initial phase of our study. It describes the skin absorption of Hz from a 70% Hz aqueous solution (H-70, used in the F-16 EPU) and from anhydrous Hz. Both anhydrous Hz and Hz from H-70 were found to be well absorbed percutaneously (86% and 55%, respectively).

This subsequent report describes the percutaneous absorption of Hz in the rabbit when exposures are terminated at various times (time-limited), and from more dilute aqueous solutions, 15% and 2%, than had been used before. The purpose of this research was to obtain additional data for the purpose of predicting the rate and extent of Hz absorption from anhydrous Hz, H-70, or more dilute aqueous Hz solutions. These additional data may also be used to test the reliability of predictions regarding percutaneous Hz absorption made in the initial report. Those predictions of bioavailability and absorption rate were made by assuming evaporation and percutaneous absorption of Hz from anhydrous Hz and H-70 applied to skin to be competing, first order processes (Keller et al., 1981).

METHODS AND MATERIALS

This study was accomplished in three parts: part one, reported previously (Keller et al., 1981), in which one group of rabbits received H₂ intravenously, and two groups were percutaneously treated with H₂ either as anhydrous H₂ or H-70; part two, in which three groups of rabbits were percutaneously exposed to anhydrous H₂ and three groups to H-70 and exposures terminated at two, six, or twelve minutes post application (time-limited exposures); and, part three, in which two groups of rabbits were percutaneously exposed to aqueous solutions of either 15% or 2% H₂.

Anhydrous H₂ (95%, Eastman, Rochester, NY) was used for neat H₂ administration and to prepare all aqueous H₂ solutions except the 15% solution. This solution was obtained as Amerzine, a catalyzed 15% aqueous solution of H₂ (Drew Chemical Corporation, Boonton, NJ). The Amerzine was tested to verify percent H₂ in solution using a spectrophotometric technique described by Reynolds and Thomas (1964). All solutions except Amerzine were prepared just prior to administration to the rabbits.

All groups of rabbits, consisting of four or five New Zealand White rabbits (Willoughby's Rabbitry, Sabina, Ohio) weighing 2.9 to 3.9 kg, received 12 mg H₂/kg. The method of animal preparation and administration of H₂ or H₂ solutions was identical to that of part one (Keller et al., 1981). Briefly, this method consisted of placing a catheter in the femoral vein, clipping and then shaving an area of skin, applying the H₂ or H₂ solution to the shaved area, and taking serial blood samples which were analyzed for serum H₂ concentration as described by Reynolds and Thomas (1964).

In part two, the percutaneous exposure to anhydrous H₂ or H-70 was terminated at the appropriate time by flushing the treated area with dilute hypochlorite solution followed by a water rinse.

The group mean serum H₂ concentrations were determined for each time period and serum H₂ versus time was graphed. In addition, the area under the serum concentration time curve (AUC) was determined for each treatment group. The percent H₂ dose absorbed for each treatment group was then determined by the equation, % dose = AUC percutaneous/AUC iv. The AUC was determined gravimetrically. The AUC iv and other data from the intravenous treatment group had been determined previously (Keller et al., 1981).

The rabbits were sacrificed 24 hours postexposure and selected hydrazine exposed skin sections were collected and examined histopathologically to determine any significant differences in skin damage with respect to H₂ solution strength and/or time of exposure.

RESULTS

Time-limitation of percutaneous H₂ exposure demonstrated a direct correlation between duration of exposure and peak serum H₂ concentrations for both anhydrous H₂ and H-70 H₂ (figures 1 and 2). A correlation of

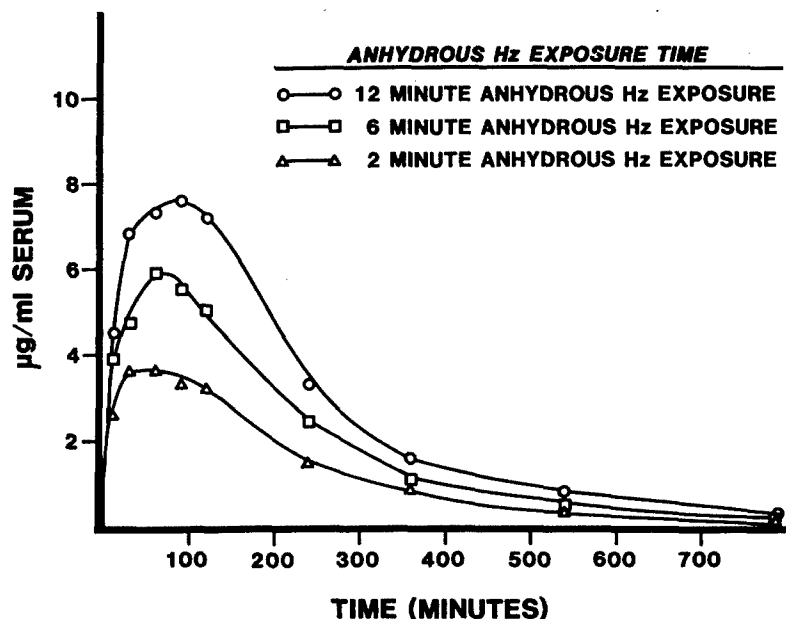
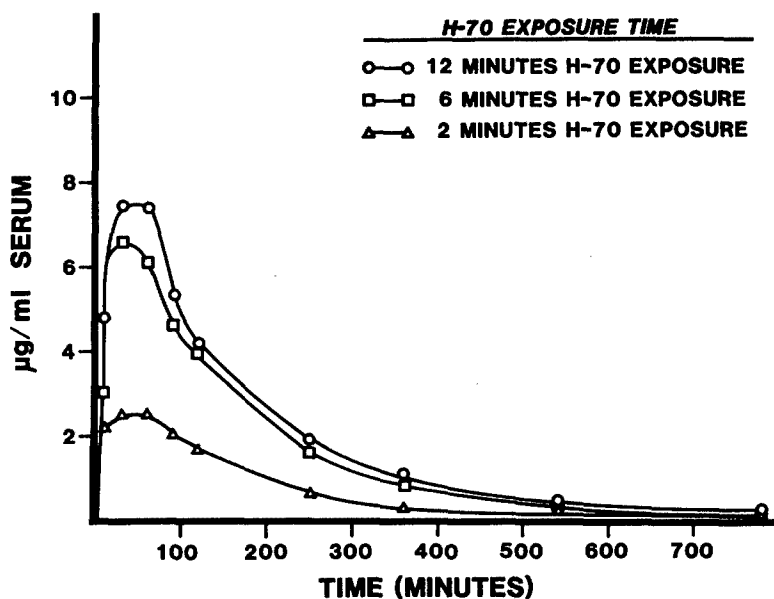


Figure 1. Serum concentration - time curves for 2, 6, and 12 minute skin exposures to anhydrous H₂.

Figure 2. Serum concentration - time curves for 2, 6, and 12 minute skin exposures to H-70.



time to peak serum H₂ level with duration of H₂ exposure was also evident with anhydrous H₂ but not H-70 H₂. Serum H₂ levels peaked at 50-90 minutes for anhydrous H₂ (Figure 1) and at 30-50 minutes for H-70 H₂ (Figure 2). Total percutaneous H₂ absorption from anhydrous H₂ or H-70 as determined by percent

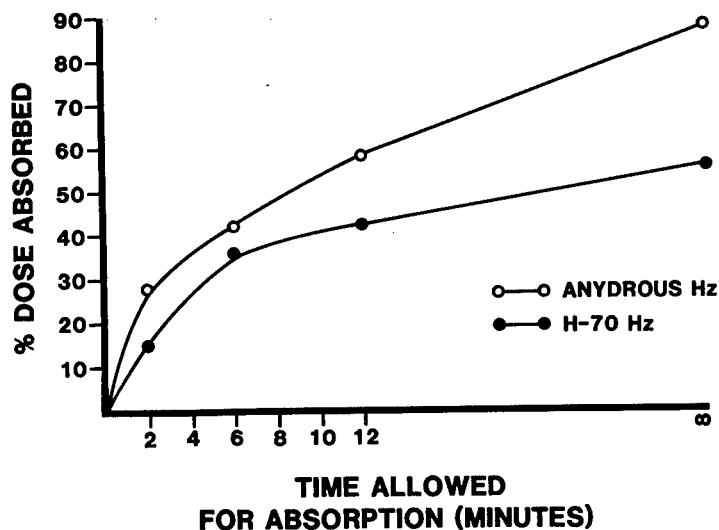
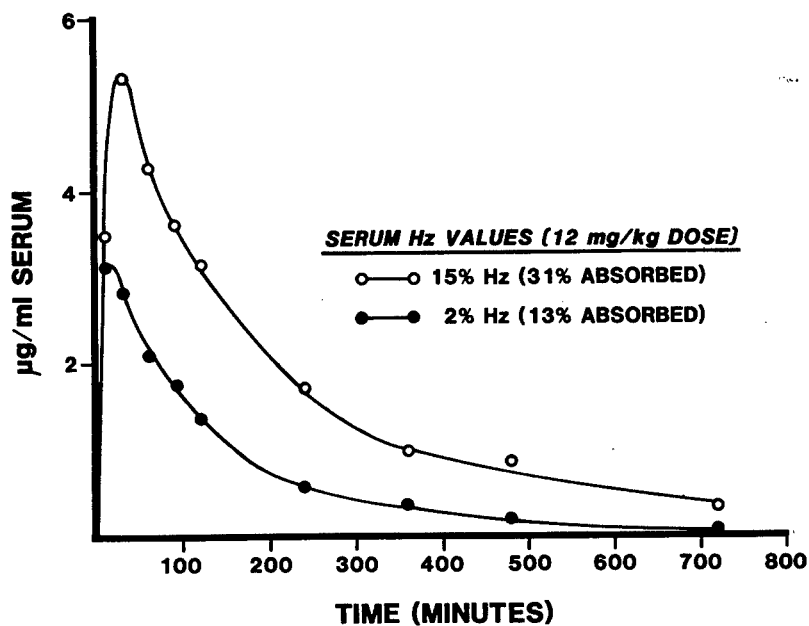


Figure 3. Percent dose absorbed versus time allowed for skin absorption of Hz and H-70.

Figure 4. Serum concentration-time curves for skin exposure to 2% and 15% Hz.



dose absorbed was also directly related to duration of Hz exposure (Figure 3). In addition, the rate of Hz absorption was greater for anhydrous Hz than for H-70 (Figure 3). The peak serum Hz concentrations for percutaneous exposure to 15% Hz and 2% Hz were 5.6 ug/ml and 3.1 ug/ml, respectively (Figure 4). These peak serum Hz concentrations occurred at 30 minutes for 15%

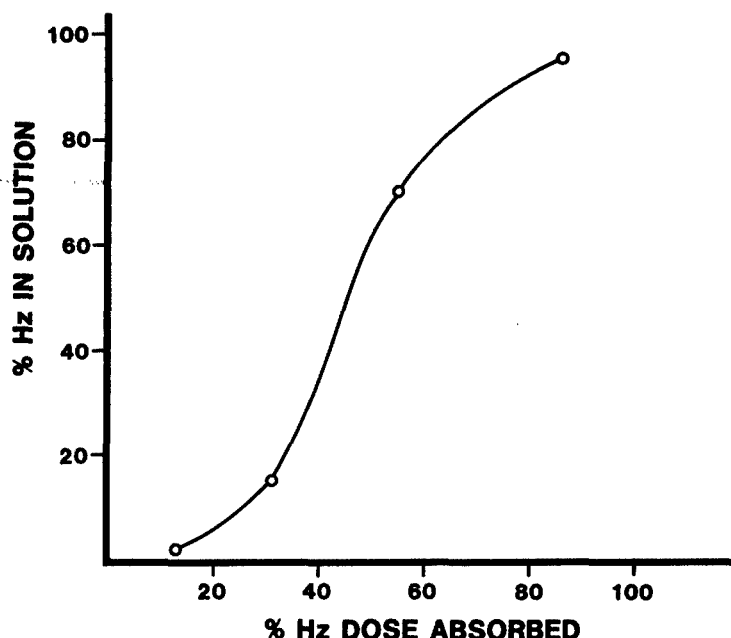


Figure 5. Hg solution concentration versus absorbed dose after skin exposure.

Hg and 10 minutes for 2% Hg. There was an increase in the percent Hg dose absorbed as the Hg solution concentration increased (Figure 5).

Severe chemical burns were found in rabbits percutaneously exposed to anhydrous Hg or H-70. These lesions typically contained areas of transepidermal necrosis with varying degrees of dermal necrosis. No significant lesions were noted following percutaneous exposure to 15% or 2% Hg solution.

DISCUSSION

A direct relationship was found between duration of percutaneous Hg contact and both peak blood concentrations of Hg and percent Hg dose absorbed through the skin for both anhydrous Hg and H-70 Hg (Figures 1, 2 and 3). These results clearly indicate that percutaneous Hg absorption, whether from anhydrous Hg or H-70 Hg, can be reduced by limiting the duration of exposure. Although the 12 minute absorption time yielded similar peak concentrations for H-70 and anhydrous Hg, the 6 minute absorption interval had a higher Hg serum concentration for H-70 and the 2 minute absorption interval had a higher Hg serum concentration for anhydrous Hg. It is apparent that the serum Hg - time curves derived from percutaneous absorption of anhydrous Hg are much broader, with a slower rate of decline, than the H-70 serum Hg - time curves. Thus, the AUC for serum Hg - time curve from anhydrous Hg absorption are larger and the percent doses absorbed are correspondingly larger (Figure 3). Also apparent (figures 1 and 2) is the time lag between percutaneous exposure to Hg and entry of Hg into the blood. All serum concentration versus time curves clearly show that blood Hg concentrations continue to increase long after Hg exposure has been discontinued. This phenomenon is more apparent with anhydrous Hg than with H-70 Hg. Since external Hg has been removed, this occurrence can only be explained by the existence of an epidermal compartment to which Hg moves from the external skin surface and from which it leaves to move into the blood. Thus, this second experiment

does seem to support the prediction made in the initial report that percutaneous H₂ absorption is best explained by including an epidermal compartment. In addition, it is also possible the function of this epidermal compartment may be more complex with anhydrous H₂ than with H-70 H₂ due to the more intense interaction of H₂ and tissue occurring with this more concentrated and caustic H₂ solution. A faster movement of H₂ from skin surface into the epidermal compartment followed by a slower but more extensive absorption into the blood would explain the broader serum H₂ - time curves from anhydrous H₂ skin exposures. Further support for the existence of an epidermal H₂ compartment can be found by inspecting figure 3. The percent dose absorbed at each time interval is greater for anhydrous H₂ than for H-70 H₂. In addition, the percent dose absorbed at 6 minutes was 36% for H-70 H₂ and 42% for anhydrous H₂. Extrapolation to five minutes gives estimates of 30% for H-70 H₂ and 39% for anhydrous H₂. This seems to suggest that the simple model of competing first order systems of evaporation and percutaneous absorption used in the initial report was not adequate for predicting anhydrous H₂ percutaneous absorption, but was more adequate for predicting the absorption of H₂ from H-70 (30% actual vs 24% predicted). In addition, these data suggest that models based solely on the rate of appearance of materials in the blood, rather than on the actual percent dose absorbed during time-limited studies, may be misleading with respect to prediction of rates of percutaneous penetration due to failure to take into account the epidermal compartment. By determining only the blood concentration - time curve, only the rate-limiting step for percutaneous absorption will be evident. In those instances when epidermal penetration rate is greater than distribution from the site of penetration (an epidermal compartment situation), a time limited study would seem to be the best approach for discerning the actual rate-limiting step of percutaneous absorption and the existence of an epidermal compartment. It would seem that the rate constants for percutaneous absorption of H₂ from H70 and anhydrous H₂ determined in our initial report are in fact rate constants for movement of H₂ from the epidermal compartment to blood rather than the rate constants for H₂ penetration of the stratum corneum. It remains to be seen whether this H₂ epidermal compartment is the result of reversible epidermal binding, creating a temporary reservoir of H₂, or simply a function of difference between rate of H₂ stratum corneum penetration and H₂ diffusion from the local skin area into the blood. The requirement for an accurate estimate of the availability of an epidermal compartment for a percutaneous agent goes far beyond the academic desire to provide a complete toxicokinetic description for an agent. Time between exposure and peak blood levels such as those described for hydrazine are in fact similar to those involved in snake envenomation. The potential for effective local treatment to minimize H₂ absorption is quite evident. Examination of data in Table 1 will allow estimates of the epidermal H₂ compartment to be made. By assuming the total bioavailable H₂ to be 100% and subtracting from it the percentage of H₂ dose absorbed at time t divided by total bioavailable H₂ absorbed, an estimate of the extent of the H₂ epidermal compartment can be made (for H-70 at two minutes: $15\%/55\% = 27\%$ and $100 - 27 = 73\%$ of bioavailable dose is available to the epidermal compartment). Thus 2/3-3/4 of the bioavailable percutaneous dose is still available for local treatment at two minutes.

Table 1

Percentage of Absorbed (Available) Dose Entering Epidermal
Compartment Following Percutaneous Application of
12 mg H₂/kg as H-70 or Anhydrous H₂

Time Interval (Minutes)	<u>H-70 (55% Total Absorption)</u>		<u>H₂ (87% Total Absorption)</u>	
	<u>% Of Total Absorbed</u>	<u>% Remaining</u>	<u>% Of Total Absorbed</u>	<u>%Remaining</u>
0-2	27	73	32	68
2-6	38	35	16	52
6-12	11	24	19	33
12-120	24	0	33	0

Further inspection of Table 1 shows an apparent comparative slowing of percutaneous absorption for anhydrous H₂ at 2-6 minutes. A physiologic explanation for this slower appearance of H₂ in the blood during percutaneous absorption of H₂ due to application of anhydrous H₂ versus H-70 H₂ may be found by examining the vasculature of the skin. Skin contains 3 levels of blood supply: a deep plexus, a middle plexus, and a superficial plexus of arteries with corresponding venous return. In addition, the lymph capillaries arise in the superficial dermis and drain into a subcutaneous plexus (Dellmann and Brown, 1981). Damage to the superficial and middle portions of skin by the more caustic anhydrous H₂ could delay movement of H₂ into the blood because the H₂ would have to diffuse through the tissue to deeper venules, and perhaps a greater portion would also be cleared by the lymphatic system, which has a much slower circulation than blood.

The absorption of H₂ from more dilute solutions has not been previously reported (Figure 5). No apparent local effect on the skin was visible following application of these dilute solutions of H₂ as was reported for more concentrated H₂ (Keller et al., 1981). Thus the absorption cannot be attributed to epidermal damage. This is a significant finding since many percutaneous occupational exposures to H₂ are to more dilute solutions of H₂ or perhaps even to H₂ vapor. These data clearly demonstrate, however, that epidermal damage is not necessary for considerable percutaneous absorption to occur.

CONCLUSIONS

Our ability to confidently make recommendations regarding the hazard of human H₂ exposures based on this study is limited by several factors: 1. Although we have corroborating data of a toxic effect following percutaneous H₂ exposure (embryo lethality in the rat, Keller et al., 1982) in another

species, no quantitative toxicokinetic data regarding percutaneous H₂ absorption are available in a species other than the rabbit; 2. The number of animals involved in this study (4-5/group) is limited; and 3. Serum H₂ was quantitated using a spectrophotometric technique which is unable to distinguish between H₂ and monoacetylhydrazine, a known metabolite, leading to a "mixed" kinetic description. Despite these limitations, some recommendations can be made particularly regarding research, which will allow a more confident estimate of the percutaneous H₂ hazard. Because of the finding of significant percutaneous absorption of H₂ from dilute solutions, further work is needed to define the hazard of percutaneous absorption of H₂ in dilute solutions or vapors and define the need for protective measures in various H₂ work environments to enhance worker protection and efficiency. Because of the delay in appearance of H₂ in the blood following percutaneous exposure, further work to define the effectiveness of local treatment may prove highly beneficial for developing techniques to treat H₂ exposures. An adjunct to both of the above may be biological sampling of H₂ - exposed workers via urine collection and analysis (Hershey et al.) as modified by George and Llewellyn (unpublished).¹ Following establishment of baseline data on H₂ in urine of exposed workers, sampling of this type might prove quite valuable for determining worker's actual daily H₂ exposure as well as for diagnostic and prognostic tests for individuals with extensive H₂ exposure.

¹ George M. E. and Llewellyn B. M., 1983. Improved Gas Chromatographic Urine Hydrazine Determination (Unpublished), Air Force Aerospace Medical Research Laboratory, Wright-Patterson Air Force Base, Ohio.

REFERENCES

Dellman, H. O. and Brown E. R., 1981. Textbook of Veterinary Histology, 2nd ed., Lea and Febiger, Philadelphia PA.

Hershey M. K., Sherman P. L., Pustinger J. V., 1979. Development of analytical techniques for toxicokinetic research, AFAMRL-TR-79-89 (AD A077 729) Air Force Aerospace Medical Research Laboratory, Wright-Patterson Air Force Base, Ohio.

Keller W. C., Murphy J. P. F., Andersen M. E., and Back K. C., 1981. Percutaneous toxicokinetics of hydrazine and H-70 in the rabbit AFAMRL-TR-81-13 (AD A097 954) Air Force Aerospace Medical Research Laboratory, Wright-Patterson Air Force Base, Ohio.

Keller W. C., Olson C. T., Back K. C., and Gaworski C. L., 1982. Evaluation of the embryotoxicity of hydrazine in rats, AFAMRL-TR-82-29 (AD A119 706) Air Force Aerospace Medical Research Laboratory, Wright-Patterson Air Force Base, Ohio.

National Institute of Occupational Safety and Health, 1978. Occupational Exposure to Hydrazine, DHEW (NIOSH) No. 78-172.

Reynolds B. A. and Thomas A. A., 1964. Determination of hydrazine and 1-methylhydrazine in blood serum, AMRL-TR-64-24 (AD 441 231) Air Force Aerospace Medical Research Laboratory, Wright-Patterson Air Force Base, Ohio.

Rothberg S. and Cope O. B., 1955. Toxicity studies on hydrazine, monomethylhydrazine, symmetrical dimethylhydrazine, unsymmetrical dimethylhydrazine, and dimethylnitrosamine, Chemical Warfare Laboratories Report No. 2027. Army Chemical Center, Edgewood, Maryland, US Army Chemical Corps Research and Development Command, Chemical Warfare Laboratories, Toxicology Division.

Smith E. B. and Clark O. A., 1972. Absorption of hydrazine through canine skin, Toxicology and Applied Pharmacology, 21, 186-193.